
Does Plasma Sequestration Reduce Post-Operative Bleeding?

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ABSTRACT

Hematologic profiles, postoperative blood loss and transfusion requirements were studied in patients undergoing cardiac surgery with cardiopulmonary bypass (CPB) in order to determine the effect of sequestration and reinfusion of platelet-rich plasma (PRP). Eighteen patients were randomly selected to have approximately one unit of PRP collected or not prior to CPB. Autologous PRP was reinfused in nine patients after reversal of anticoagulation. Evaluation of results was based upon thrombocyte counts, Hgb, Hct, and platelet index readings pre- and post-PRP reinfusion, amount of blood loss and volumes of homologous blood and banked blood products required postoperatively. There were no hemodynamic complications related to the sequestration process. PRP reinfused patients had significantly higher thrombocyte counts after reversal of anticoagulation ($p < 0.05$). Patients receiving PRP required approximately 2/3 less banked blood products, their bleeding being significantly reduced 24 hours postoperatively ($p < 0.05$). We concluded that plasma sequestration and reinfusion of autologous PRP post-bypass may serve as an effective and safe way in decreasing blood loss after cardiac operations necessitating extracorporeal circulation.

INTRODUCTION

Patients undergoing cardiac surgeries with cardiopulmonary bypass (CPB) are prone to developing serious hemorrhagic diathesis. Incomplete neutralization of heparin,^{1,2} decreased plasma coagulation factors,^{3,4} and excessive fibrinolysis,³⁻⁶ remain some of the major derangements seen after CPB. Platelet activation and consumption during CPB, causing postoperative thrombocytopenia as well as platelet dysfunction, are recognized as strong factors which induce bleeding after cardiac surgery.^{8,9} Methods for preparation and storage of homologous platelets are available, however, their randomly or selectively matched transfusion for correction of post-bypass

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bleeding is not without risks. Complications of platelet transfusions are similar to those of red cells. In addition, alloimmunization to histocompatibility antigens and refractoriness to random donor platelets may occur.¹⁰ Transfusion-transmitted diseases,^{11,12} bacterial contamination,¹³ non-hemolytic transfusion reactions,¹⁴ graft-versus-host disease¹⁵ and significant cost are only some of the potential problems related to platelet replacement from banked sources. In the era whereby concerns that demand of blood products may exceed the supply, interest in finding ways to counteract platelet disturbances after CPB has increased. Used initially in community blood centers to collect plasma, the Haemonetics Plasma Saver System has evolved as an instrument to collect Platelet-Rich Plasma (PRP) prior to CPB. When reinfused after anticoagulant neutralization, PRP may contain important clotting elements lost during CPB, their replacement resulting in improved hemostasis and a significant reduction of blood loss.

We undertook this study to establish the effects of platelet sequestration and reinfusion of PRP on hematologic parameters, postoperative blood loss and transfusion requirements following cardiac surgery with extracorporeal circulation.

MATERIALS AND METHODS

This randomized prospective study was performed on 18 adult patients undergoing elective cardiac surgeries at Cooper Hospital/University Medical Center in Camden, New Jersey. The patients included in this study did not take any antiplatelet drugs for at least one week prior to their operation. All patients received standard anesthesia induced with fentanyl, etomidate and pancuronium, maintained with oxygen and nitrous oxide, and supplemented with fentanyl. Ventilation with 100% oxygen was adjusted to maintain normocarbida. Standard CPB was instituted in all patients using a roller pump^a with PVC tubing. Heparin (3mg/kg) was used prior to cannulation and activated clotting time (ACT) was frequently checked to measure the level of anticoagulation. ACT was maintained over 480 seconds during CPB by supplemental heparin. CPB was conducted under moderate hypothermia. Non-pulsatile flow between 2.1 L (hypothermia)

and 2.4 L/min./m (normothermia) was employed. The extracorporeal circuit was primed with 2 liters of Ringer's injection^b and 150 cc of 25% albumin.^c One ampule of NaHCO₃ and 5,000 units of heparin were also added. A membrane oxygenator^d was employed in all cases.

All patients signed a consent form to participate in the study, which was approved by the Institutional Review Board. In nine randomly selected patients, Haemonetics Plasma Saver System^e was used to remove PRP. After induction of anesthesia, the amount of PRP removed was in accordance with the patient's weight, body build and hematocrit level. Blood was drawn from a large gauge internal jugular catheter.^f Fluid was given prior to starting the collection of PRP and sufficient volume replacement (lactated Ringer's solution) was infused during the collection to replenish the lost volume. Haemonetics Plasma Saver System is composed of a blood pump, centrifuge assembly programmable to variable rates, red cell detector, anticoagulant pump, plasma weigher, four air detectors and a sensor to monitor the blood pressure. In accordance with the manufacturer's instructions, the actual centrifuge rate used in this study was 3000 rpm. Centrifugation of the PRP at 4000 rpm for 10 minutes results in platelet injury, while, 3000 rpm maintains platelet viability. PRP was collected, properly labeled, placed on a "rocker" and stored at room temperature. Blood pressure was regulated with sodium nitroprusside.

Upon completion of CPB, heparin was antagonized with protamine sulfate (1:1 with heparin), half given as bolus and half as a continuous infusion. When ACT returned to, or near baseline level, PRP was reinfused to the patient. Blood samples for hematological measurements were collected by venipuncture preoperatively (baseline), during CPB at hypothermia (28°C) and normothermia (34°C), 15 minutes upon termination of CPB (after PRP was reinfused), and at 24 and 48 hours postoperatively. Hematological assessments consisted of platelet counts, hemoglobin and hematocrit measurements and were assessed by standard laboratory techniques. A platelet index was calculated by dividing the platelet count by the hematocrit value. Postoperative blood loss was estimated by the volume of blood present in the chest drainage reservoir from the time of chest tube insertion intraoperatively through the first 48 hours. The content of the oxygenator was returned to each patient at the end of the procedure as centrifuged blood. Transfusion amounts of banked whole blood, packed red cells (PRC) and fresh-frozen plasma (FFP) were also recorded. No intra- or post-operative deaths occurred. All patients had an uneventful hospital course. In both groups, reoperation for bleeding was not necessary. SPSS PC Plus (version 2.0) was used for statistical analysis. Paired t-test and analysis of variance (ANOVA) were used where appropriate for comparison between variables at specific times. Values are presented as mean standard deviation. A p value of < 0.05 was considered statistically significant.

a. COBE Laboratories, Lakewood, CO

b. and c. Travenol Laboratories, Deerfield, IL

d. COBE Laboratories, Lakewood, CO

e. Haemonetics Corporation, Braintree, MA

f. Arrow International, Reading, PA

RESULTS

The demographic and hematologic profiles were similar between the two groups (Tables 1 and 3). The intraoperative conditions, including the time on CPB and the surgical procedures performed, were also comparable (Tables 2 and 3). During CPB, hypertension was controlled with sodium nitroprusside or phenylephrine and mean arterial pressure was maintained between 60 and 85 mm Hg. There were no major hemodynamic reactions to jeopardize or interrupt the collection of PRP. In both groups, CPB produced a statistically significant reduction in platelet count, Hct. and Hgb. level (Table 1). Significantly low hematological values were observed up to 48 hours postoperatively. In the control group, the platelet count decreased further after reversal of heparin with protamine sulfate (post-bypass); this was not seen in the PRP-reinfused group ($p < 0.05$). Twenty-four and 48 hours after surgery, platelet counts in the two groups were not statistically different (122.8 ± 70.9 vs 127.8 ± 31.5 and 128.4 ± 36.8 vs 131.6 ± 30.2 respectively, $p > 0.05$). At all times, comparison of the platelet index between the two groups did not reach statistical significance (Table 1). Comparable amounts of centrifuged blood retrieved from the oxygenator circuit were reinfused in both groups ($p > 0.05$). The plasma sequestered group received a mean of 220.8 ± 61 ml PRP (range 121 to 300 ml.) representing 9% of the total platelet volume. The control group received none. However, there was a striking difference in blood loss between the control and PRP-reinfused group over the first 24 hours (630.0 ± 78.5 vs 407.8 ± 87.7 ml., $p < 0.05$). During the second postoperative day the differences in blood loss between the two groups did not reach statistical significance. However, PRP-reinfused patients had 25% less total blood loss at the end of 48 hours postoperatively (963.3 ± 80.5 vs 718.9 ± 99.0 , $p < 0.05$). The need for transfusion of banked blood products was significantly different. Patients in the control group received 65% more blood products given as FFP and PRC than those receiving autologous PRP (total 24 units of FFP plus 15 units of PRC vs four units of FFP plus 12 units of PRC); 9,750 ml of blood products were used in the control group and only 3,500 ml were used in the PRP-reinfused group ($p < 0.05$).

DISCUSSION

The hemostatic alterations observed after CPB, explained in part by the hemodilution or displacement of the coagulation fibrinolysis balance toward a tendency of hemorrhage, is especially considered to be the effect of real depletion of the coagulation cascade due to trauma to the blood components and plasma denaturation. Triggered by the contact of blood elements with the extracorporeal circuit, this process may result in disseminated intravascular coagulation (DIC) and the activation of vasoactive kinins and complement systems¹⁶⁻¹⁸. Activation of the Hageman Factor XII commonly initiates the body's stress reaction system (coagulation, fibrinolysis, complement and kine activation).¹⁹ CPB produces thrombocytopenia which persists for several days post-operatively.^{8,20} A loss of platelet function characterized by increases in plasma thromboxane B^{21,22} plasma, beta-

TABLE 1
CHARACTERISTICS OF PLASMA SEQUESTERED GROUP VS. CONTROL

Characteristics	Base Line	Bypass		
		Cold	Warm	
		(28°C)	(34°C)	
Platelets Count (x10³)				
Control Group	253.7 ± 65.9	118.3 ± 38.1	139.3 ± 46.2	
PS Group	235.2 ± 64.1	93.1 ± 27.9	129.9 ± 36.5	
Significance**	NS	NS	NS	
Hematocrit (%)				
Control Group	33.2 ± 4.3	19.0 ± 2.8	20.1 ± 2.6	
PS Group	34.3 ± 4.2	19.7 ± 2.5	21.7 ± 2.5	
Significance**	NS	NS	NS	
Hemoglobin (g/dl)				
Control Group	11.4 ± 1.5	6.5 ± 0.8	6.8 ± 0.9	
PS Group	11.5 ± 1.5	6.7 ± 0.8	7.3 ± 0.7	
Significance**	NS	NS	NS	
Platelet Index (x10³)				
Control Group	7.8 ± 2.3	5.9 ± 1.6	6.9 ± 1.9	
PS Group	6.9 ± 2.1	4.3 ± 0.5	6.0 ± 1.3	
Significance**	NS	NS	NS	
Characteristics	Post Bypass (15 min.)	Postoperative Day 1	Significance* Day 2	
Platelets Count (x10³)				
Control Group	95.4 ± 15.6	122.8 ± 70.9	128.4 ± 36.8	p < 0.05
PS Group	135.7 ± 28.1	127.8 ± 31.5	131.6 ± 30.2	p < 0.05
Significance**	p < 0.05	NS	NS	
Hematocrit (%)				
Control Group	22.5 ± 4.4	25.8 ± 3.2	27.8 ± 3.3	p < 0.05
PS Group	22.3 ± 1.9	26.4 ± 1.2	26.6 ± 3.0	p < 0.05
Significance**	NS	NS	NS	
Hemoglobin (g/dl)				
Control Group	7.5 ± 1.3	8.8 ± 1.0	9.5 ± 0.8	p < 0.05
PS Group	7.3 ± 0.6	8.9 ± 0.6	9.1 ± 0.9	p < 0.05
Significance**	NS	NS	NS	
Platelet Index (x10³)				
Control Group	5.4 ± 1.1	6.3 ± 2.3	4.7 ± 1.6	
PS Group	6.0 ± 0.8	4.8 ± 1.1	4.2 ± 1.3	
Significance**	NS	NS	NS	

PS = plasma sequestered; All values are mean ± standard deviation;
NS = not significant *ANOVA = between values in the same group
S**= t-test between mean values of the 2 groups

TABLE 2
TYPE OF OPERATIONS PERFORMED

TYPE OF OPERATION	NO. OF PATIENTS	
	CONTROL (%)	PLASMA SEQUESTRATION (%)
CABG	5 (55.6%)	4 (44.5%)
Valve Replacement (VR)	2 (22.2%)	3 (33.3%)
Combined Procedures*	2 (22.2%)	2 (22.2%)
TOTALS	9	9

CABG - Coronary Artery Bypass Grafting

* CABG plus VR

TABLE 3
GENERAL CHARACTERISTICS

Variable	Control (n=9)	PS (n=9)	Significance
AGE (yr)	58.2 ± 3.4	59.0 ± 1.8	NS
BSA (m ²)	1.9 ± 0.9	1.9 ± 0.2	NS
Perfusion Time (min.)	128.4 ± 4.5	125.8 ± 6.8	NS
Blood Loss (ml)			
Postop Day 1 (0-24 hrs.)	630.0 ± 78.5	407.8 ± 87.7	p < 0.05
Postop Day 2 (24-48 hrs.)	344.4 ± 72.3	311.1 ± 37.6	NS
TOTALS	963.3 ± 80.5	718.9 ± 99.0	p < 0.05
Blood Received			
Centrifugated Blood (ml)	525.0 ± 50.0	542.0 ± 35.0	NS
Autologous PRP (ml)	-----	220.8 ± 61.0	
Bank Blood (units)	24 FFP + 15 PRC	4 FFP + 12 PRC	p < 0.05
Total Bank Blood (ml)	9,750	3,500	p < 0.05

Values are presented as mean ± standard deviation

NS = Not Significant

PS = Plasma Sequestered Group

thromboglobulin and platelet factor IV,^{8,21} and decreases in platelet beta-thromboglobulin and fibrinogen will occur.⁴ Additionally, selective loss of alpha-granules,⁸ reduction of thrombocyte aggregability,^{3,23,24} and their adhesiveness²⁵ and increased bleeding time⁸ have been extensively related to extracorporeal circulation. Also, neutralization of heparin with protamine sulfate at the completion of CPB produces a damaging dose-dependent effect on platelets.²⁶⁻²⁸ In order to reverse some of the derangements seen after CPB and to try to restore clotting factors to levels required for normal hemostatic response, homologous banked blood products have been extensively used.^{8,29-32} There are, however, potential complications related to crossmatching errors, allergic reactions, isoimmunization and contamination. Also a significant reduction of blood banked products have been signaled during recent years.

This study was initiated to assess the efficacy and safety of a commercially available Plasma Saving System. The Haemonetics Plasma Saver System has the capability to sequester a certain amount of PRP prior to the initiation of CPB and to reinfuse it after completion of heparin neutralization and to return it into the circulation as non-altered, viable thrombocytes dispersed into physiologically normal plasma. Our study demonstrated that sequestration of 9% of the total platelet volume does not produce any additional significant alterations of the hematologic course during CPB. Reinfusion of autologous PRP significantly increased thrombocyte count levels post-bypass, a time when further alteration in coagulation profile induced by heparin neutralization with protamine sulfate may eventually produce diffuse bleeding. There are several possible reasons for improved platelet function after administration of autologous PRP. First, PRP contains thrombocytes which are not damaged by storage as are those in banked platelet concentrates.³³⁻³⁵ Second, fibrinogen, factor V and VIII (von Willebrand) which are decreased during CPB, and are inactive or destroyed in banked blood products³⁶ but fully active in freshly collected PRP, may be responsible for the better hemostatic effect of reinfused-PRP. Improvement in the post-bypass platelet counts with normal functional quality made possible a significant reduction in post-CPB bleeding at the end of 24 and/or 48 hours. In our study, patients receiving autologous PRP required 65% less homologous blood to control postoperative bleeding, thereby being exposed to considerably less risk of transfusion-transmitted diseases. In summary, our observations demonstrated that reinfusion of autologous PRP may significantly reduce the need for homologous blood requirement following CPB in a safe and tolerable manner.

REFERENCES

1. Soloway HB, Cornett, BM, Donahoo et.al.: Differentiation of bleeding diathesis which occur following protamine correction of heparin anticoagulation. *Am J Clin Path.* 59:188-91, 1973.
2. Lambert CJ, Marengo-Row AJ, Leveson J, et.al.: The Tri-F titer: A rapid test for estimation of plasma fibrinogen and detection of fibrinolysis, fibrin(ogen) split products, and heparin. *Ann Thorac Surg.* 18:357-63, 1974.
3. Mammen EF, Koets MH, Washington B, et.al.: Hemostasis

- changes during cardiopulmonary bypass surgery. *Sem Thromb Hemostas.* 11:281-92,1985.
4. Mezzano D, Aranda E, Urzua J, et.al.: Changes in platelet beta-thromboglobulin, fibrinogen, albumin, 5-hydroxytryptamine, ATP, and ADP during and after surgery with extracorporeal circulation in man. *Am J Hematol.* 22:133-42, 1986.
 5. Muller N, Popov-Cenic S, Buttner W, et.al.: Studies of fibrinolytic and coagulation factors during open-heart surgery. II. Postoperative bleeding tendency and changes in the coagulation system. *Thromb Res.* 7:589-98, 1975.
 6. Lambert CJ, Marengo-Row AJ, Leveson JE, et.al.: The treatment of post-perfusion bleeding using epsilon-aminocaproic acid, cryoprecipitate, fresh frozen plasma, and protamine sulfate. *Ann Thorac Surg.* 28:440-4, 1978.
 7. Holloway DS, Summaria L, Sandesara J, et.al.: Decreased platelet number and function and increased fibrinolysis contribute to postoperative bleeding in cardiopulmonary bypass patients. *Thromb Haemostas.* 59(1): 1988.
 8. Harker LA, Malpass TW, Branson HE, et.al.: Mechanism of abnormal bleeding in patients undergoing cardiopulmonary bypass: Acquired transient platelet dysfunction associated with selective alpha-granule release. *Blood.* 56:824-34, 1980.
 9. McKenna R, Brachmann F, Whittaker B, et.al.: The hemostatic mechanism after open-heart surgery. II. Frequency of abnormal platelet functions during and after extracorporeal circulation. *J. Thorac Cardiovasc Surg.* 70:298-308, 1975.
 10. Dahlke MB, Weiss KL.: Platelet transfusion from donors mismatched for cross reaction HLA antigens. *Transfusion.* 24:299-302, 1984.
 11. Anderson KC, Gorgone JBC, Lew M.: Transfusion related sepsis after prolonged platelet storage. *Blood.* 66 (Suppl I):273a, 1985.
 12. Braine HG, Kichler RS, Charache P, et.al.: Bacterial sepsis secondary to platelet transfusion: An adverse effect of extended storage at room temperature. *Transfusion.* 26:391-3, 1986.
 13. Gorgone BC, Lew M, Anderson DC.: Bacterial contamination of cytopheresis platelets (Abstract). *Transfusion.* 26:584, 1986.
 14. Buck SA, Kickler TS, Braine HG, et.al.: Elimination of allergic platelet transfusion reactions by automated platelet washing (Abstract). *Transfusion.* 25:459, 1985.
 12. Leitman SF.: Post-transfusion graft-versus-host disease. In: Smith, D.M., Silvergleid, A.J. eds. Special considerations in transfusing the immunocompromised patient. Washington, D.C.:AABB. 15-37, 1985.
 16. Lindsay RM.: Blood surface interactions (Panel Conference). *Trans Am Soc Artif Inter Organs.* 26:603, 1980.
 17. Stratta P, Canavese C, Mangiarotti G, et.al.: Heparin is unable to prevent contact activation by three different membranes. *Proc Eur Dial Transplant Assoc.* 18:269, 1981.
 18. Kirklin JK, Westaby S, Blakstone EH., et.al.: Complement and the damaging effect of cardiopulmonary bypass. *J Thorac Cardiovasc Surg.* 86:845, 1983.
 19. Stratta P, Canavese C, Costa P, et.al.: Biological stress induced by extracorporeal circulation: Comparison between

- cardiopulmonary bypass, hemodialysis and plasma exchange. *Trans Am Soc Artif Inter Organs*. 30:502-7, 1984.
20. Bachmann F, McKenna R, Cole ER, et.al.: The hemostatic mechanism after open-heart surgery. I. Studies on plasma coagulation factors and fibrinolysis in 512 patients after extracorporeal circulation. *J Thorac Cardiovasc Surg*. 70:76-85, 1975.
21. Addonizio VP, Smith JB, Strauss JF, et.al.: Thromboxane synthesis and platelet secretion during cardiopulmonary bypass with bubble oxygenator. *J Thorac Cardiovasc Surg*. 79:91-6, 1980.
22. Davies GC, Sobel M, Salzman EW.: Elevated plasma fibrinopeptide A and thromboxane B levels during cardiopulmonary bypass. *Circulation*. 61:808-14, 1980.
23. Boers M, van den Dungen JJAM, Karliczek GF., et.al.: Two-membrane oxygenators and a bubbler: A clinical comparison. *Ann Thorac Surg*. 35:455-62, 1983.
24. Boonstra PW, van Imhoff GW, Eysman L, et.al.: Reduced platelet activation and improved hemostasis after controlled cardiotomy suction during clinical membrane oxygenator perfusions. *J Thorac Cardiovasc Surg*. 89:900-6, 1985.
25. Bick RL, Arbogast N, Crawford L, et.al.: Hemostatic defects induced by cardiopulmonary bypass. *Vasc Surg*. 9:228-43, 1975.
26. van den Dungen JJAM, Karliczek GF, Brenken U, et.al.: Clinical study of blood trauma during perfusion with membrane and bubble oxygenators. *J Thorac Cardiovasc Surg*. 83:108-16, 1982.
27. Mielke CH, deLeval M, Hill JD., et.al.: Drug influence on platelet loss during extracorporeal circulation. *J Thorac Cardiovasc Surg*. 66:845-54, 1973.
28. Velders AJ, van den Dungen JJAM, Westerhof NJ, et.al.: Platelet damage by protamine administration: Protection by reducing protamine or by prostacyclin (PG12) treatment. *Proc Eur Soc Artif Organs*. 6:194-7, 1979.
29. Woods JE, Traswell HF, Kirklin JW, et.al.: The transfusion of platelet concentrates in patients undergoing heart surgery. *Mayo Clinic Proc* 42:318-25, 1967.
30. Harding SA, Shakoob MHM Grindon AJ.: Platelet support for cardiopulmonary bypass. *J Thorac Cardiovasc Surg*. 70:350-2, 1975.
31. Pliam MB, McGoon DC, Tarhan S.: Failure of transfusion of autologous whole blood to reduce banked blood requirements in open heart surgical patients. *J Thorac Cardiovasc Surg*. 70:338-43, 1975.
32. Mohr R, Golan M, Martinowitz U, et.al.: Effect of cardiac operation on platelets. *J Thorac Cardiovasc Surg*. 92:434-41, 1986.
33. Slichter SJ, Harker LA.: Preparation and storage of platelet concentrates. I. Factors influencing the harvest of viable platelets from whole blood. *Br J Haematol*. 34:395-9, 1976.
34. Moroff G, Friedman A, Rabkine Kline L, et.al.: Reduction of volume of stored platelet concentrates for use in neonatal patients. *Transfusion*. 24:144-6, 1984.
35. Valeri CR.: Circulation and hemostatic effectiveness of platelets stored at 4°C or 22°C: Studies in aspirin-treated normal volunteers. *Transfusion*. 16:30-3, 1976.
36. Osterud B, Rapaport SI, Lavine KK.: Factor V activity of platelets: Evidence for an activated Factor V molecule and for a platelet activator. *Blood*. 49:819-23, 1977.